

Hypothesis:
 OHC electromotility evolved from
 hair cell synaptic mechanisms.

Adventurous Motility

Edge of a *Myxococcus xanthus* colony - individual bacteria showing adventurous gliding motility, time lapse 600x speed (Kaiser lab website - Stanford).

Outer membrane ripples on motile cells:
 Coincidence or functional roles?

OHC - Dieler et al. 1991

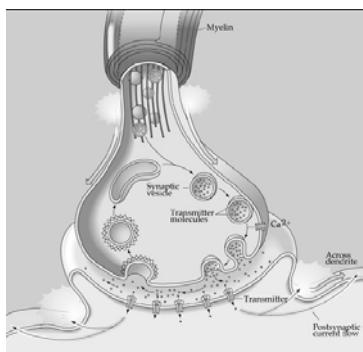
Oscillatoria - Adams et al. 1999

Flexibacter BH3 - Dickson et al. 1980

Trilaminate Walls

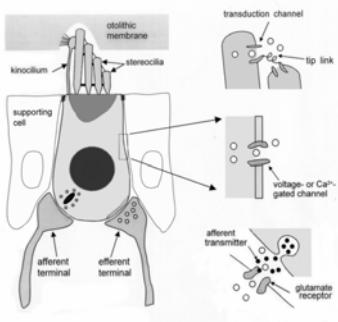
The diagram illustrates the trilaminate wall structure across three layers: OHC, Oscillatoria, and Flexibacter. Key components labeled include the outer membrane (OM), periphery (PM), cortical lattice, spectrin, actin, eosin pillar, axoneme (Ax), and the cytoplasmic plate (CP). Labels F, PG, and CM are also present.

Neural membrane curvature



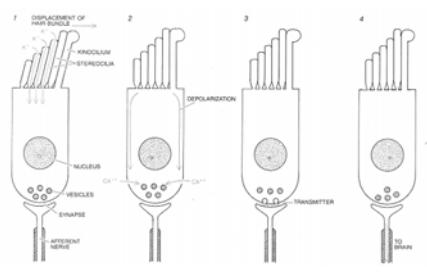
Hair Cells Have Two Functions

Mechano-Electrical Transduction



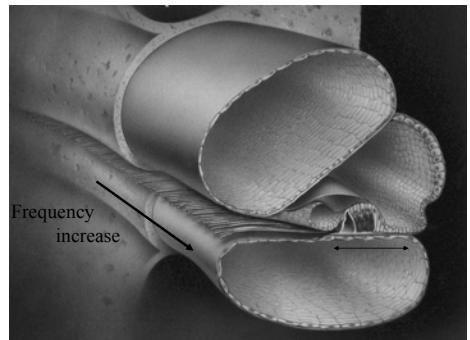
Neurotransmission

Hair Cell Neurotransmission



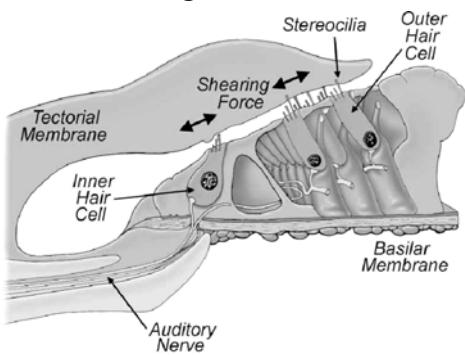
Hudspeth, 1983

The output is neural



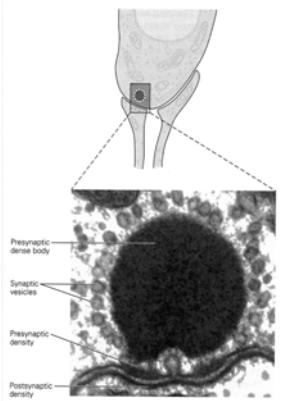
Compton's Interactive Encyclopedia, 1997

The organ of Corti

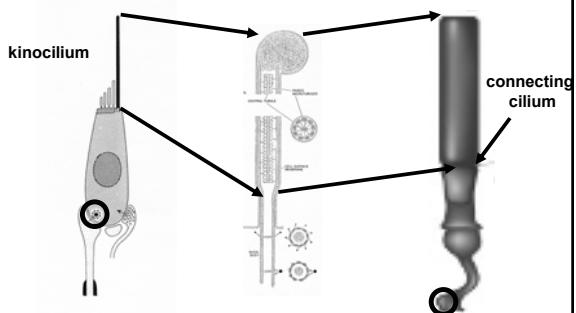


Sensitive Synapse

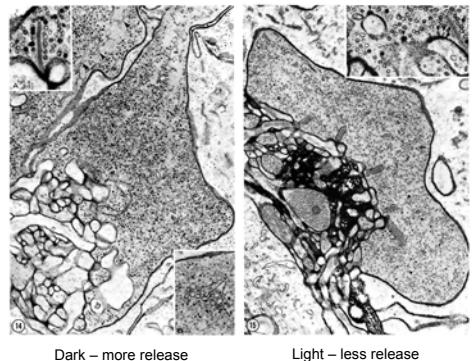
- Continuous release of neurotransmitter
- Rate of release modulated by mV changes in membrane potential



Patterns - hair cells & photoreceptors

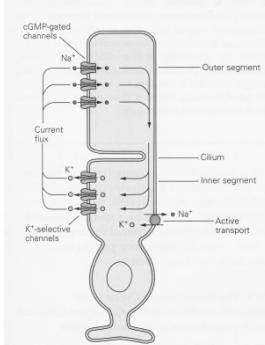


Light dependent transmitter release

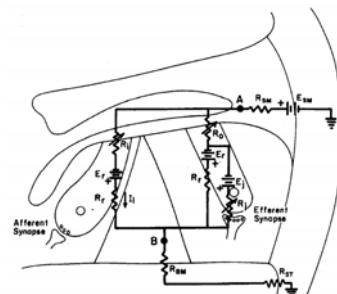


Phototransduction - dark current

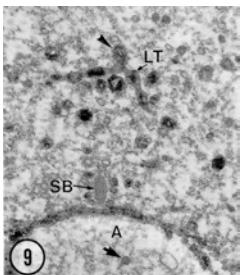
The dark current depolarizes the membrane potential resulting in maximal neurotransmitter release. Light blocks the depolarizing current and decreases neurotransmitter release.



Visualizing the silent current

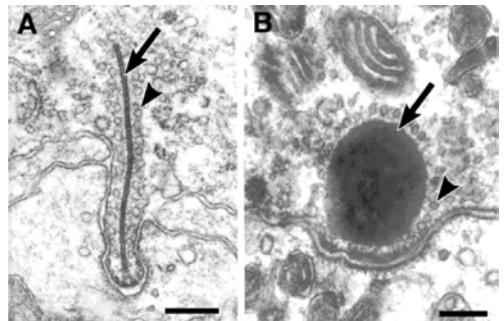


Inner hair cell afferent synapse



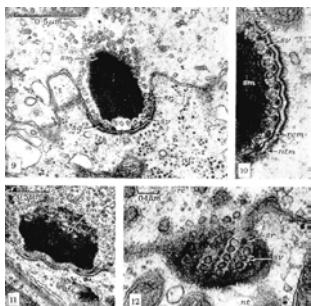
Siegel & Brownell - 1986

Ribbon synapses



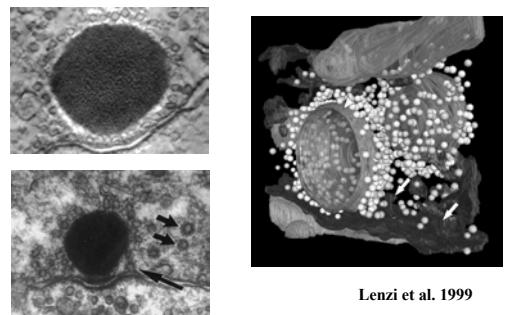
Lenzi & von Gersdorff, 2001

Ampullary organ of the North American Catfish



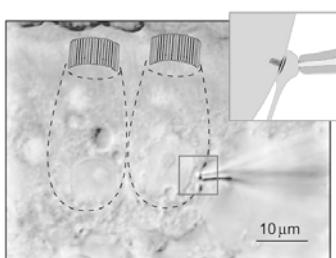
Mullinger, 1964

3-D reconstruction of frog hair cell synaptic ribbon



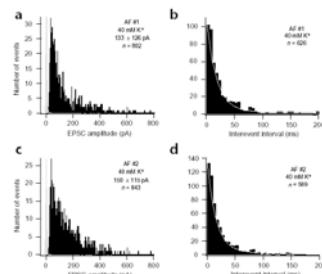
Lenzi et al. 1999

Recording from the synapse



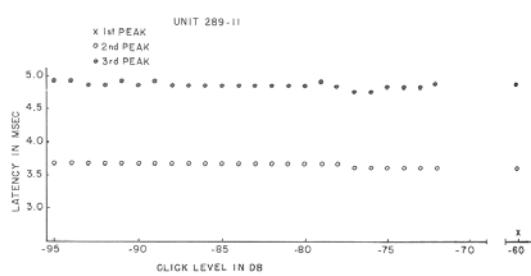
Glowatzki & Fuchs, 2002

Interval between neurotransmitter release is Poisson



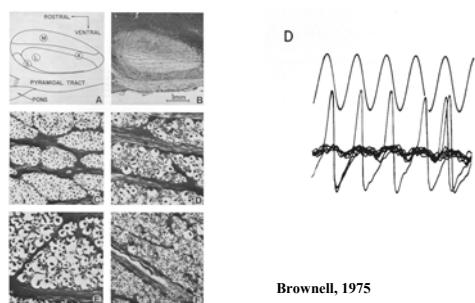
Glowatzki & Fuchs, 2002

Temporal precision



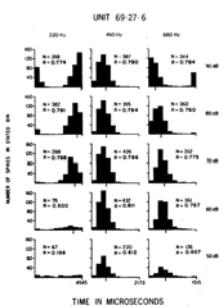
Kiang et al., 1965

Phase locking



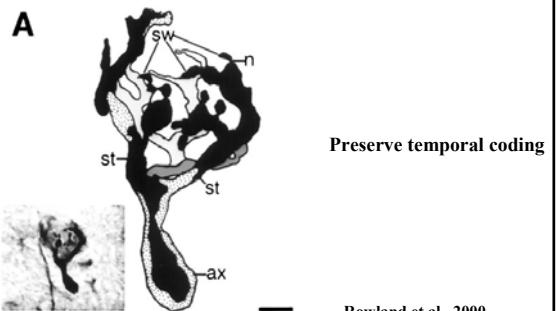
Brownell, 1975

Intensity - invariance

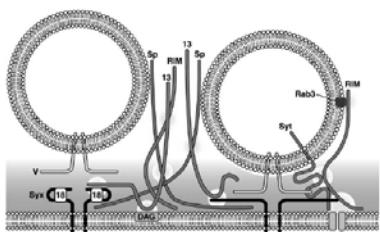


Anderson, 1971

Specialized CNS synapses

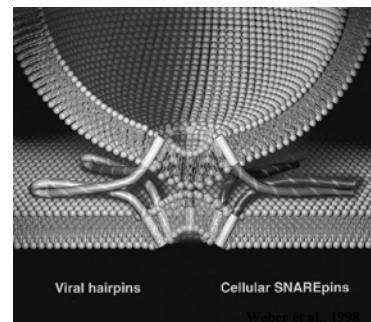


Protein-Protein Interactions in the Active Zone Matrix

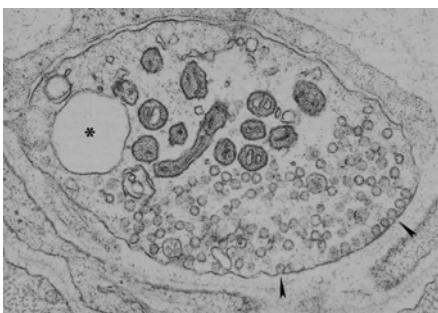


Martin, 2002

Proteins bring membranes together

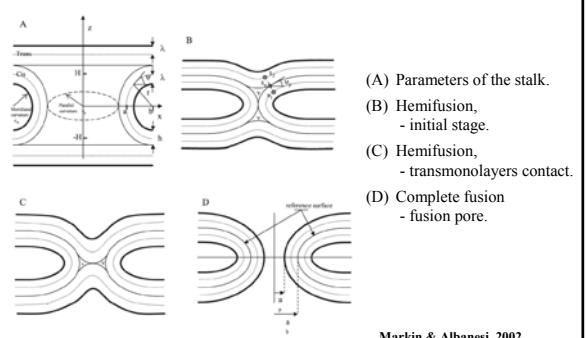


Vesicle fusion



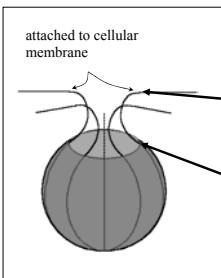
Torri-Tarelli et al., 1985

Membrane fusion



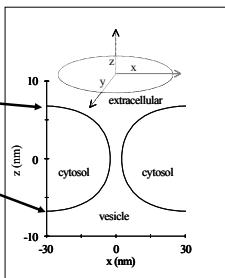
The fusion pore

Geometry

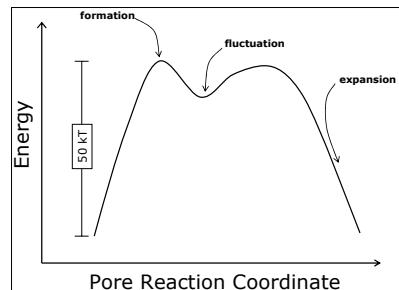


Farrell & Cox, 2002

Model



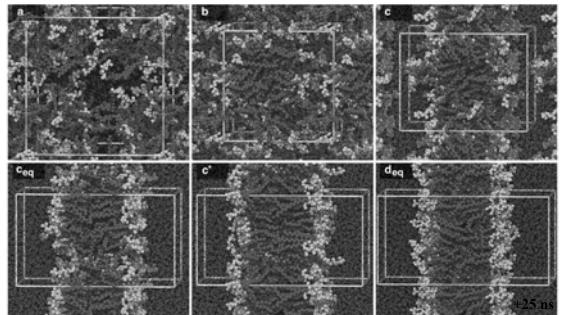
Where do we get the energy to bend the membranes?



Phospholipids: the forgotten molecules

Don't forget water

Membrane self assembly



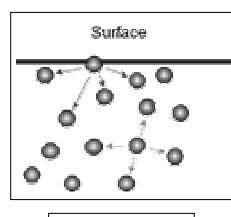
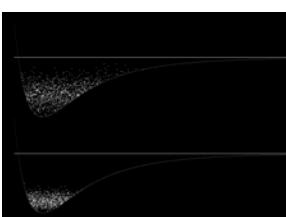
Marrink, Lindahl, Edholm & Mark, 2001



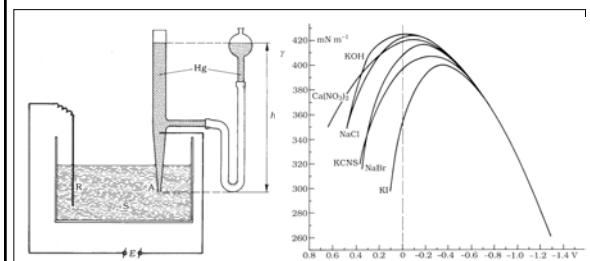
Surface tension

the energy required to increase the surface area of a liquid by a unit amount

attraction \leftrightarrow Energy \rightarrow repulsion



Electrical potential changes γ : Lippmann mercury voltmeter

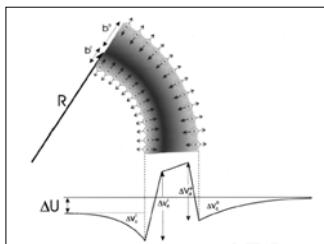


G. Lippmann, Ann. Phys. 149 (1873)

DC Grahame (1947)

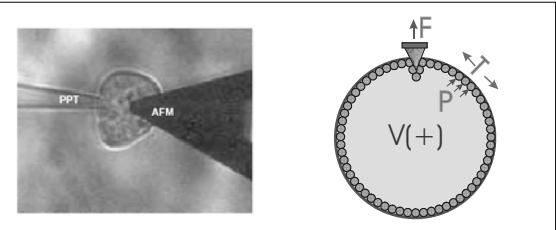
Voltage dependent membrane tension

Includes a differential change in surface tension at the two membrane interfaces



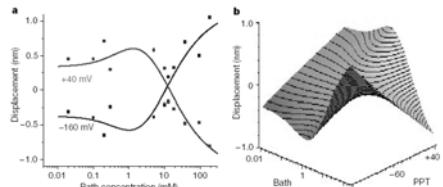
Petrov & Sachs, 2002

HEK electromotility measured under voltage clamp with AFM



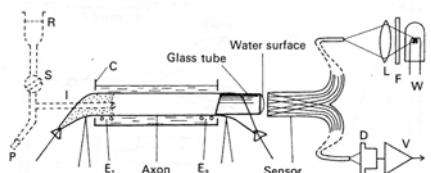
Zhang et al., 2001
Mosbacher et al., 1998

Voltage dependent membrane motion

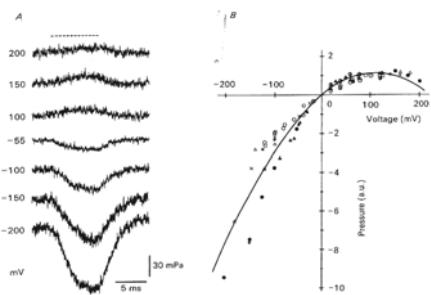


Zhang et al., 2001

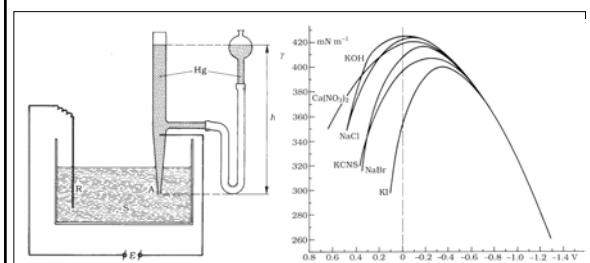
Voltage dependent pressure changes in squid axon



Terakawa, 1984



Electrical potential changes γ : Lippmann mercury voltmeter



G. Lippmann, Ann. Phys. 149 (1873)

DC Grahame (1947)

Lippmann equation

A Gibbs adsorption equation for a polarizable interface,



Gabriel Lippmann
Nobel Prize,
physics 1908

$$\sigma\gamma = -S_A \partial T - \Gamma \partial \mu - \sigma \partial E$$

contains the observed relation between surface charge and the ratio of the change in surface tension to the change in electrical potential,

$$\sigma^o = -\left(\frac{\partial \gamma}{\partial E}\right)_{(\mu, T)}$$

σ^o : surface charge (Cm^{-2})

γ : surface tension (Nm^{-1})

E : electrical potential difference (V)

μ : chemical potential (\bar{V})

T : temperature ($^{\circ}K$)

Γ : surface concentration one component (moles m^{-2})

S_A : interfacial entropy per unit area ($JK^{-1}m^{-2}$)

Voltage Dependent Tension

$$\sigma^o = -\left(\frac{\partial \gamma}{\partial E}\right)_{(\mu, T)}$$

Integrate the Lippmann under these boundary conditions:

$$1. \quad C = \epsilon \epsilon_0 \kappa_i$$

$$\kappa^2 = F^2 \sum_j \left[\frac{n_j z_j^2}{\epsilon \epsilon_0 R T} \right]$$

2. At voltage V^o

$$\sigma_i^o = -\sigma_i^d \text{ when } \gamma_i = \gamma_i(V^o) \text{ and } \sigma_e^o = -\sigma_e^d \text{ when } \gamma_e = \gamma_e(V^o)$$

Upon polarization to voltage V_m

$$\sigma_i^o = -(\sigma_i^d - q_p) \quad \gamma_i = \gamma_i(V_m) \text{ and } \sigma_e^o = -(\sigma_e^d + q_p) \text{ when } \gamma_e = \gamma_e(V_m)$$

F: Faraday's constant; n_j : concentration of species, j ; z_j : valency of species, j ; R : gas constant; ϵ_0 : permittivity of free space; ϵ_w : dielectric constant of water; C : capacitance of double layer

Tension is a linear function of voltage under physiological conditions ($\Delta V < 100$ mV)

$$T_V \approx \frac{(C_m)^2}{2} (B_e + B_i)(\Delta V)^2 + C_m(-B_e \sigma_e^o + B_i \sigma_i^o) \Delta V$$

T_V : voltage dependent tension. C_m : membrane capacitance

$$B_e = \pm \frac{1}{\kappa_e \epsilon \epsilon_0} \quad B_i = \pm \frac{1}{\kappa_i \epsilon \epsilon_0}$$

Assume:

Physiological medium 0.14 M

External and internal surface charge -0.025 and -0.015 C/m²

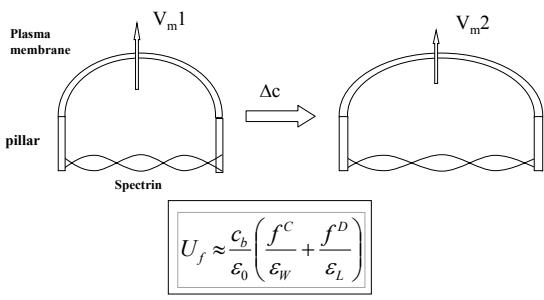
Charging occurs by ions adsorbing onto or desorbing from $\Delta V = 100$ mV

Energy \approx Pore Area * T_V

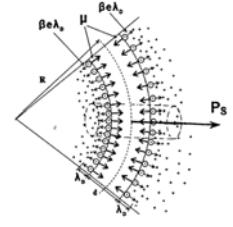
1000-5000 nm² * 46 μN/m

10-50 kT

The flexoelectric effect



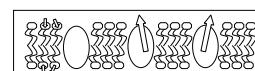
Flexoelectricity: coupling of membrane curvature with the electric field



characterized in biological membranes
by Petrov

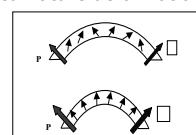
Liquid Crystal Nature of Biomembranes

Protein and lipid molecules comprising biomembranes possess dipole moments



Dipoles contribute to the flexoelectric effect

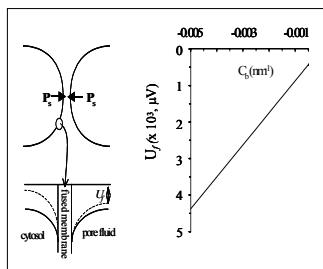
- curvature deformation changes membrane polarization



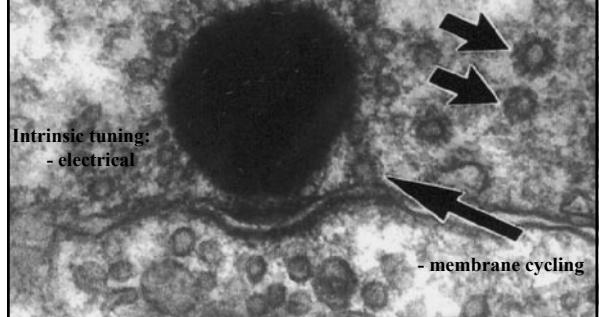
As c is increased, dipoles become more aligned increasing the polarization of the membrane

Direct flexoelectric effect

$$U_f \approx \frac{c_b}{\epsilon_0} \left(\frac{f^C}{\epsilon_w} + \frac{f^D}{\epsilon_L} \right)$$



The synaptic amplifier



OHC electromotility – the other membrane based motor



Collaborators

Baylor:

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Brenda Farrell, Ph.D.
Olivier Lichtarge, M.D., Ph.D.
Fred Pierera, Ph.D.
Peter Saggau, Ph.D.

Buffalo:

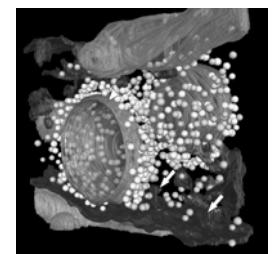
Fred Sachs, Ph.D.
Ken Snyder

Johns Hopkins:

Aleksander S. Popel, Ph.D.
Alexander A. Spector, Ph.D.

Rice:

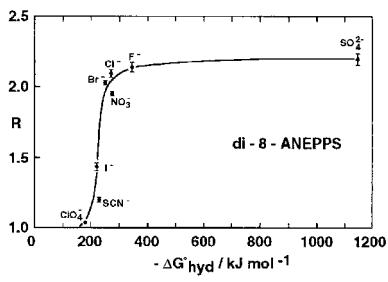
Bahman Anvari, Ph.D.
Robert M. Raphael, Ph.D.



URLs: <http://www.bcm.tmc.edu/oto/research/cochlea/>
<http://www.bioflexoelectricity.org>

Hofmeister effect

anion adsorption at membrane interface

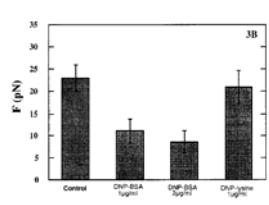
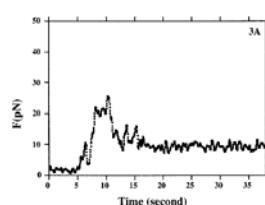


$\text{ClO}_4^- > \text{SCN}^- > \text{I}^- > \text{NO}_3^- > \text{Br}^- > \text{Cl}^- > \text{F}^- > \text{SO}_4^{2-}$

$\text{I}^- > \text{Br}^- > \text{NO}_3^- > \text{Cl}^- > \text{HCO}_3^- > \text{F}^- > \text{SO}_4^{2-}$

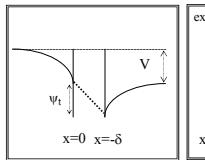
Oliver et al., 2001

Tension affects vesicle recycling

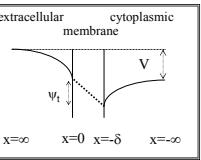


Dai et al., 1997

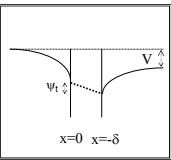
Hyperpolarization & depolarization affect the charge on each interface



Hyperpolarization



Rest



Depolarization

ψ_t : potential difference across the membrane
 V : transmembrane potential difference

$$\psi_t = \psi(-\delta) - \psi(0)$$

$$V = \psi(-\infty) - \psi(\infty) = \psi(-\infty)$$