

A blue-tinted background image of a microscope. The objective lenses are visible, with some text like '8' and '10' on them. The text is overlaid in a dark red color.

# Human Dermal Fibroblast Cell Proliferation and Viability in Vitro

YYY

BIOE 342

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# Experimental Objectives

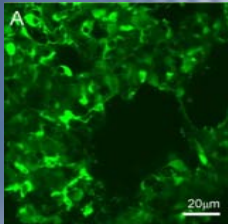
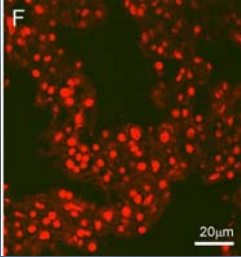
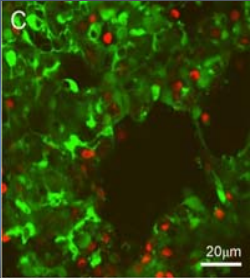
- To assess qualitatively Human Dermal Fibroblast (HDF) viability with fluorescence imaging
  - Live/Dead Fluorescence Assay
- To measure quantitatively the effects of serum concentration on HDF growth and proliferation
  - Anti-PCNA Staining Assay
  - Cell Proliferation Assay

# Live/Dead Fluorescence Assay

- Preparation: HDF cells seeded in 24-well TC treated plates
  - Incubated 2 days
  - Dyed with solution of Calcein AM and EthD-1
- Conditions (3 wells each):
  - A. Control
  - B. 1 mL Ethanol
  - C. 2 drops Ethanol
- Equipment: Fluorescent microscope



# Dyed Cells Fluoresce Green When Alive and Red When Dead

Condition	Example <sup>1</sup>	Observations
Dye alone	 Fluorescence micrograph A shows a field of cells stained green. The cells are elongated and appear to be attached to a surface. A scale bar in the bottom right corner indicates 20 μm.	All cells are stained green. Cells are elongated and attached.
Dye, 1 mL Ethanol	 Fluorescence micrograph F shows a field of cells where the nuclei are stained red. The cells are mostly rounded. A scale bar in the bottom right corner indicates 20 μm.	All cell nuclei are stained red. Cells are mostly rounded.
Dye, 2 drops Ethanol	 Fluorescence micrograph C shows a field of cells with a mix of green and red staining. Some cells are green, while others have red nuclei. The cells are a mix of elongated and rounded shapes. A scale bar in the bottom right corner indicates 20 μm.	Some green cells interspersed with red nuclei. Some elongated and rounded cells. There are more green cells than red stained cells.

# HDF Cells Are Permeable to Ethanol

- HDF membranes allow the diffusion of ethanol into the cell
- Ethanol is disruptive to HDF cell environment and results in visible cell necrosis
- Small amounts of ethanol kill only a portion of the HDF cells
- We may use live/dead assays to assess the cytotoxicity of any other substance

# Anti-PCNA Staining Assay

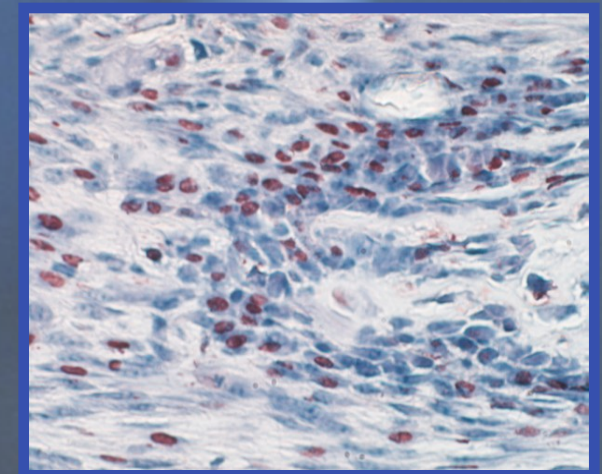
- Preparation: Cells seeded at ~20,000 cells/mL in 24-well TC treated plates
  - Incubated 2 days
  - Fixed in Formalin
  - Primary Antibody (Ab): Anti-PCNA Mouse IgG
  - Secondary Ab: Anti-mouse IgG tagged with HRP
- Conditions (1 well each):
  - Dulbecco's Modified Eagle Medium (DMEM) with 1, 5 and 10% Fetal Bovine Serum (FBS)
  - In addition, 3 controls seeded with 10% FBS DMEM
- Equipment: Light microscope



# Fraction of HDF in S Increases with %FBS

- HDF in S phase are preparing for division
  - PCNA production at a max in S phase
- PCNA tagged by primary and secondary Ab
  - HDF cells in S phase stain red<sup>2</sup>
- Percentage of HDF in S phase increases 85% from 1 to 5 % FBS, and 74% from 1 to 10% FBS

Condition	Confluence	Cell Staining Observations
1% FBS	60%	50% blue, 50% red
5% FBS	60-80%	7.2% blue, 92.8% red, many double nuclei visible
10% FBS	80%	13% blue, 87% red, double nuclei visible

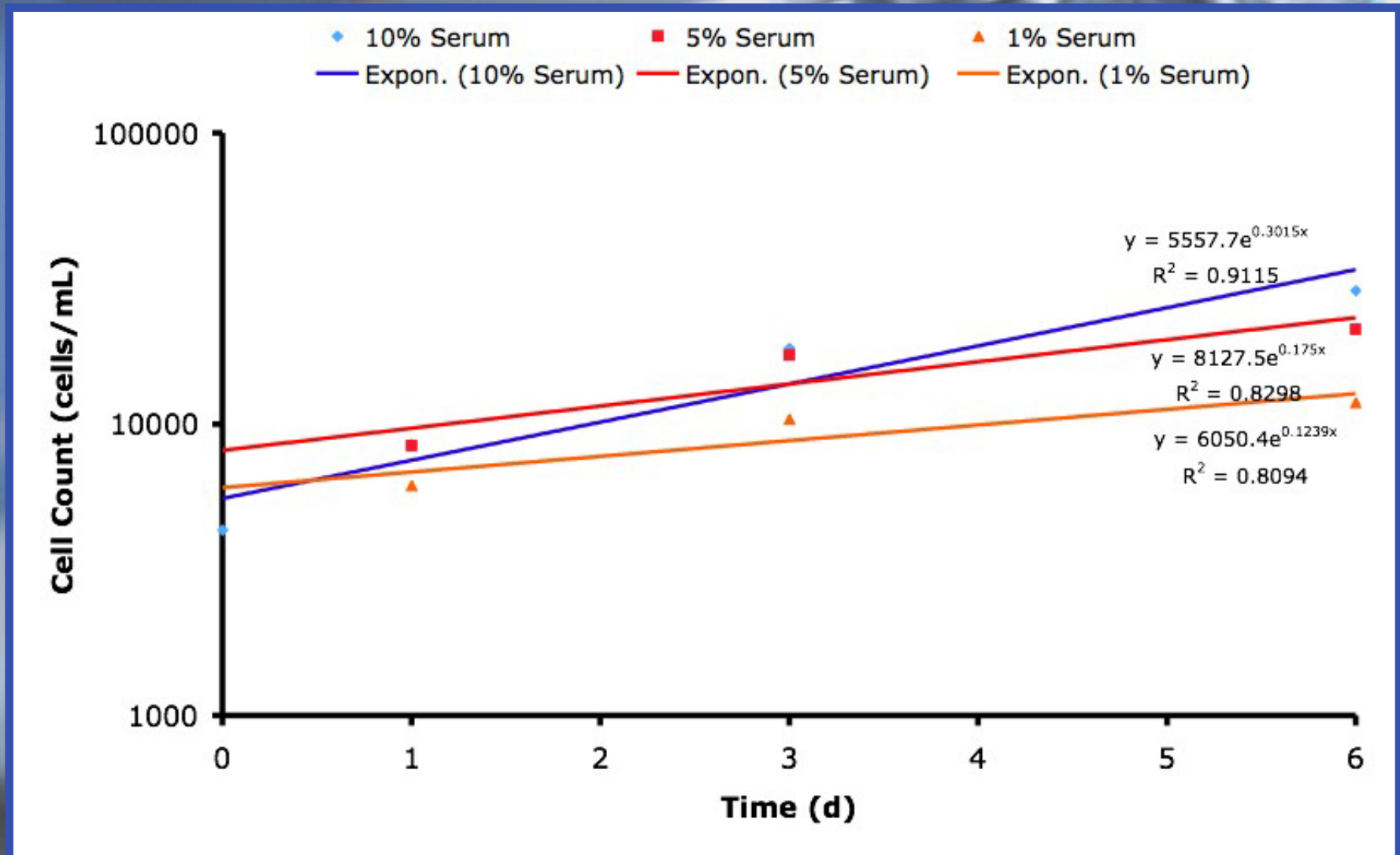


# HDF Proliferation Assay

- Preparation: Cells seeded at ~5000 cells/mL in 24-well TC treated plates
  - Media replenished on day 1
- Conditions (Data obtained at 0,1,3 and 6 days):
  - DMEM with 1,5 and 10% FBS
- Equipment:
  - Light microscope
  - Coulter Counter



# HDF Growth Curve is Exponential and Related to %FBS



# 10% FBS Yields Higher Cell Count Than 1% and 5%

- As % FBS increases from 1 -10%, [cell] increases exponentially ( $R^2 > 0.8$ )
- Day 3 cell count means among 1, 5 and 10% FBS are statistically different as determined by ANOVA with Tukey's HSD ( $p = 0.0447 < 0.05$ )
- Doubling times decrease with increasing %FBS

Coulter Counter Data (Day 3)

1% FBS	5% FBS	10% FBS
682	935	648
350	871	1075
433	785	1015

Serum (%)	Time to double (d)
10%	2.3
5%	3.96
1%	5.59

# Proliferation and Anti-PCNA Results Show Same Trend

- Both assays illustrate HDF cells' propensity towards FBS in media
- Increasing serum concentration from 1 to 10% results in:
  - Increased final (day 6) cell concentration by 74%
  - Decreased doubling time by 41%
  - Larger proportion of cells in S phase (More viable proliferating cells) by 85%
- FBS nourishes cells with nutrients and growth factors
  - Increasing its concentration in media is strongly advised at least up to 10%



# Key Results

- Cytotoxicity may be assessed using fluorescence imaging as in the Live/Dead assay
  - Ethanol decreases cell viability by quickly disrupting the cell environment
  - Qualitative data: morphology, spread, distribution
  - Visualization is subject to human error
- Proliferation assays provide quantitative data from which to derive the exponential growth curve relationship as well as doubling times
  - Varies with %FBS
- Anti-PCNA assays show the fraction of cells that are likely proliferating
  - Log phase of the growth curve
  - Varies with %FBS

# Work Cited

- 1
  - <http://respiratory-research.com/content/figures/1465-9921-6-40-1-l.jpg>
- 2
  - [http://www.cor.uams.edu/images/youngmouse\\_do%20pcna40x.jpg](http://www.cor.uams.edu/images/youngmouse_do%20pcna40x.jpg)